### 2015

# M.Sc. Part-I Examination ZOOLOGY

PAPER-II (Group-A)

Full Marks: 50

Time : 2 Hours

The figures in the margin indicate full marks.

Candidates are required to give their answers in their own words as far as practicable.

Illustrate the answers wherever necessary.

### Group-A

Answer any four questions taking two from each unit.

## Unit—I Cytogenetics |

1. (a) A cross is made between an Hfr that is met thr pur and an F that is met thi pur. Interruptied mating studies show that met enters the recipient last, so that met recombinants in the F back ground are selected on a medium containing supplements that

satisfy only the pur and thi requirements. These recombinants are tested for the presence of the thi and pur alleles. The following numbers of individuals are found for each genotype:

- 1. met thi pur 280
- 2. met thi pur 0
- 3. met thi pur 6
- 4. met thi pur 52
- (a) Why was methionine (met) left out of the selection medium?
- (b) What is the gene order?
- (c) What are the map distances in the recombination  $2\frac{1}{2}+5+5$ units?
- 2. (a) Various pairs of r II mutants of phage T4 are tested in E.coli in both cis and trans positions. Comparisons are made of 'burst size'. Table shows results for six different r mutants: rM, rN, rO, rP, rR, and rS. If we assign the mutaion ro, to the A cistron, what are the locations of the other five mutations with respect to A and B cistron?

Result of r II mutant crosses

Cis genotype	Burst size	Trans genotype	Bust size	
rM rN/++	245	rM+/rN	250	
rO rP/++	256	rO+/+rP	268	
rR rS/++	248	rR+/+rS	242	
rM rO/++	270	rM+/+rO	0	
rM rP/++	255	rM+/+rP	255	
rM rR/++	264	rM+/+rR	0	
rM rS/++	240	rM+/+rS	240	
rN rO/++	257	rN+/+rO	268	
rN rP/++	250	rN+/+rP	0	
rN rR/++	245	rN+/+rR	255	
rN rS/++	259	rN+/+rS	0	
rP rR/++	260	rP+/+rR	245	
rP rS/++	253	rP+/+rS	0	

(b) The following deletion map shows four deletions (1-4) involving the rII A cistron of phage T4:

4.	-				
2.				14.4	
		DI LEAFE I	ST CONTRACT	357 . 18	
0					

Five point mutations (a-e) are tested against these tone deletion mutants for their ability to give wild type (r<sup>+</sup>) recombinants, the results are:

What is the order of the point mutation?

3. (a)		AA	Aa	aa
.0200	1	0.3	0.0	0.7
80000	II	0.2	0.2	0.6

Which of the above populations are in HW-equilibrium?

- (b) About 70% all white North American can taste phanyl-thiocarbamide & the remainder can not. The ability to taste is determined by the dominant allele T, and the inability to taste is determined by the recessive allele. If the population is assumed to be in HW equilibrium, what are the genotypic and allelic frequencis in this propulation?
- (c) In a generalized transducing phage P.1, the donor is pur nad pdx and the receipient is pur nad pdx.

  The donor allele pur is initially selected after transduction, and 50 pur transductants are then scored for the other alleles present. The result follow:

Genotype	Number of colonies
nad pdx	Infunction 3
nad pdx	and or grafion at rach
nad pdx +	24
nad pdx	ie structus out shots 13 notiseller notisellere
morgalizative si	et netraelles des 3 an

- (i) What is the cotransduction frequency for pur & nad?
- (ii) What is the contransduction frequency for pur & pdx?
  - (iii) Which of the unselected loci is closest to pur?  $3+3+6\frac{1}{5}$
- (a) Mention the role of Ras protein in a signaling cascade with proper diagram.
  - (b) What happens when a cell containing two mutant rb alleles?
  - (c) How does  $G_1$  arrest take place when DNA damage occurs?  $4+4+4\frac{1}{2}$

#### Unit-II

### [Molecular Biology]

- 5. (a) What is the key to the high processivity of the DNA polymerase?
  - (b) State the features of the "trombone" model for coordinating replication by two DNA polymerases at the *E.coli* replication fork with diagram.
  - (c) What is meant by polymerase switching?

 $^{4+5+3\frac{1}{2}}$ 

- 6. (a) How does termination occur in E.coli translation?
  - (b) State the role of elongation factor, EF G in E.coli.
  - (c) What is the role of initiation factor IF 1 and IF 3 in bacteria?
- 7. (a) This question involves the lac operon of E.coli where I = repressor gene, P = promoter gene O = operator gene Z = β galactosidase gene Y = permease gene.
  Complete Table below using + to indicate that the enzyme will be synthesized and to indicate that enzyme will not be synthesized:

- (i) ISP+O+Z+Y+
- (ii) ISp+OCZ+Y-
  - (iii) I<sup>-d</sup>P<sup>+</sup>O<sup>+</sup>Z<sup>+</sup>Y<sup>+</sup>
- Assume a poly (A) tail of Z<sup>+</sup>Y<sup>+</sup>Z<sup>+</sup>O<sup>+</sup>Z<sup>+</sup>O<sup>+</sup>Z (vi) uh dots.

  I<sup>+</sup>P<sup>+</sup>O<sup>c</sup>Z<sup>+</sup>Y<sup>-</sup> Occur in the dots and dots.
- (v)  $I^{-d}P^{+}O^{+}Z^{+}Y^{+}$  $I^{+}P^{+}O^{+}Z^{-}Y^{+}$ 
  - (b) In presence of high intracelluar concentration of tryptophan, only short transcripts of the trp operon are synthesized because of attenuation of transcription to the structural genes. This is mediated by the recognition of two Trp codons in the leader sequence. What effect would mutating these two codons to UAG stop codons have on the regulation of the operon in the presence or absence of tryptophan? Explain.

8. (a) The following figure shows the transcribed region of a typical enkaryotic protein-coding gene:

Exon1	INTRON1	Exon2	INTRON2	Exon3
<b>←</b> 100 <b>←</b> :	→ ← 75 →	←50→	<del>← 70 →</del>	←25→
		DENIKE L	Frankling.	Poly (A) site

What is the size (in bases) of the fully processed, matured m RNA?

Assume a poly (A) tail of 200 as in your calculation.

- (b) How does Rho dependent termination occur in bacteria?
- (c) Match each term (1-4) with its corresponding description (s) in a-g, noting both that each term may have more than one description & each description may apply to more than one term
  - 1. Eukaryotic m RNA s
- 2. Prokaryotic m RNA s
- - 4. Ribosomal RNA s
- a. have a cloverleaf structure.
- b. are synthesized by RNA polymerases.
  - c. display one anticodon each.
- d. are the template of genetic information during translation.
  - e. contain exon & intron.
  - f. are the four types in eukaryotes & only three types in E.coli.
  - g.— are capped on their 5' end & polyadenylated on their 3' end.